

DNA extraction from Tissue (mouse tail)

Preparation of test samples

Cut 5mm of the mouse tail (c.a.10mg) and put it in the micro test tube (usually use 0.2ml or 0.5ml tubes for PCR)



Reagents

Mix the Cellease A, B and distilled water (20 ul Cellease A, 20 ul CellEase B, 60 ul distilled water)



Add 100 ul of the mixture to the samples.



Incubate at 37°C for 6 minutes
Then incubate at 95°C for 3 minutes



Transfer 6ul of extracts to PCR reaction mixture and amplify the target DNA fragment

PCR

6.0ul Test sample
5.0 ul ×10 buffer(+Mg²⁺)
5.0 ul dNTPs
1.0 ul Forward Primer (10pmol/ul)
1.0 ul Reverse Primer (10pmol/ul)
0.5 ul Ex Taq (5 U/ul)

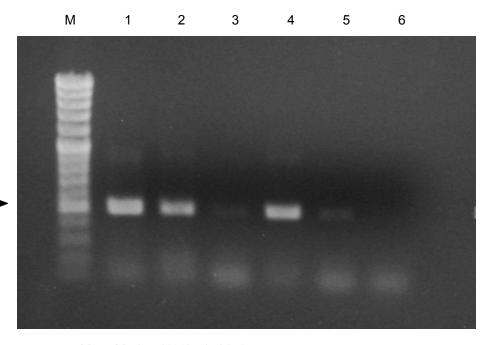
Fill up to 50 ul by distilled water

PCR Cycle

	94°C	1min	,
	94°C	30sec	35 Cycles
	55°C	30sec	
l	72°C	60sec	
	72°C	4min	

Primers: Mouse β -globin gene (494bp)

Comparison with conventional CellEase kit >



M	Marker (100bp ladder)	
1	DNA extract by using CellEase Tissue II	conc.
2		× 10
3		× 100
4	DNA extract by using conventional CellEase	conc.
5		× 10
6		× 100

- X The protocol of conventional CellEase and CellEase II kit were followed by the instruction manual respectively.
- X The thickness of the DNA bands were depending upon the amount of test samples and a parts of tissue including bone, skin or fat.

The clear DNA bands were detected from more than $\,\times\,10$ dilution of DNA extracts by using CellEase Tissue II



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